



## Short communication

## Determination of ginsenosides Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub> in rat plasma by a rapid and sensitive liquid chromatography tandem mass spectrometry method: Application in a pharmacokinetic study

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## ABSTRACT

A sensitive rapid resolution liquid chromatography–tandem mass spectrometry method was developed to determine the pharmacokinetics of ginsenoside Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub> in rats, after oral administration (50 mg/kg) and intravenous administration (10 mg/kg) of Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub>, respectively. The plasma samples were extracted by saturated N-butanol with Rg<sub>2</sub> as internal standard. Chromatographic separation was performed on a Zorbax SB-C18 column (50 mm × 4.6 mm, 1.8 μm) with a mobile phase consisting of methanol and 1 mM ammonium formate (74:26, v/v). Multiple reaction monitoring mode was performed using the fragmentation transitions of  $m/z$  1107.7 →  $m/z$  178.9,  $m/z$  1077.7 →  $m/z$  148.6, and  $m/z$  1077.7 →  $m/z$  783.4 for Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub>, respectively. Calibration curves were recovered over a concentration range of 20–1000 ng/ml for Rb<sub>1</sub> and Rb<sub>2</sub>, and 50–2500 ng/ml for Rb<sub>3</sub>. The limits of detection were 3.0 ng/ml, 4.0 ng/ml, and 6.5 ng/ml. Both intra-day and inter-day variances were less than 15% and the accuracy was within 86–114% for the three ginsenosides. All three ginsenosides had poor oral bioavailability (0.78%, 0.08%, and 0.52% for Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub>, respectively). The value of Rb<sub>1</sub> is higher than that of Rb<sub>2</sub> or Rb<sub>3</sub>, indicating that ginsenosides with hexose and hydroxyl groups (Rb<sub>1</sub>) could present better pharmacokinetic behaviors than those with pentose groups in the same glycosylation site by oral administration.

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## 1. Introduction

Ginsenosides have been regarded as the main active components responsible for the pharmacological activities of *Panax* herbs [1]. Ginsenoside Rb<sub>1</sub> is the main saponin with Rb<sub>2</sub> and Rb<sub>3</sub> as the other minor chief constituents in *Panax* extract. The three ginsenosides all are 20(S)-protopanaxadiol saponins (PPD) and have the same glycosylation site (C-3 site, C-20 site). The difference only lies in glycoside connected to the C-20 site. Rb<sub>1</sub> connects with two glucoses, Rb<sub>2</sub> connects with one glucose and one α-L-arabinopyranosyl, and Rb<sub>3</sub> connects with one glucose and one β-D-xylose (Fig. 1).

Until now, the pharmacokinetic characteristics of ginsenosides have been published by many literatures. Liu et al. found that Rb<sub>1</sub>, Rb<sub>2</sub>, Rc and Rd were all distributed rapidly in rat brain, muscle and nervous tissues after intravenous administration [2]. Song

et al. reported a pharmacokinetic study of R<sub>1</sub>, Rg<sub>1</sub> and Rb<sub>1</sub> in dogs after oral administration. The results showed the C<sub>max</sub> for Rb<sub>1</sub> was 28.6 ng/ml [3]. Chen et al. reported that the t<sub>1/2</sub> of Rb<sub>1</sub> is much longer than Rg<sub>1</sub> and R<sub>1</sub> [4]. The bioavailability of Rb<sub>1</sub> has been reported to be 4.35% [5]. The study on the absorption and disposition of Ra<sub>3</sub>, Rb<sub>1</sub>, Rd, Re, Rg<sub>1</sub>, R<sub>1</sub> showed that most ginsenosides were subject to rapid extensive biliary excretion, resulting in their short t<sub>1/2</sub> values [6].

Although the pharmacokinetic characteristic of Rb<sub>1</sub> is known, the information of Rb<sub>2</sub> and Rb<sub>3</sub> are rare. Actually, limited information has been published on pharmacokinetic behaviors of Rb<sub>2</sub> and Rb<sub>3</sub>. Therefore, pharmacokinetic studies of Rb<sub>2</sub> and Rb<sub>3</sub> are necessary and could help clarify the relationship between pharmacokinetics and structural preference of ginsenosides. Since ginsenosides have poor UV radiation in maximum absorption of 203 nm [5]. The liquid chromatography mass spectrometry (LC-MS) [7–9] and liquid chromatography–tandem mass spectrometry (LC-MS/MS) [10–12] method are widely used for analyzing ginsenosides.

In this study we developed a simple, rapid, and sensitive LC-MS/MS method to determine plasma concentrations of Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub> after oral and intravenous administration of the three

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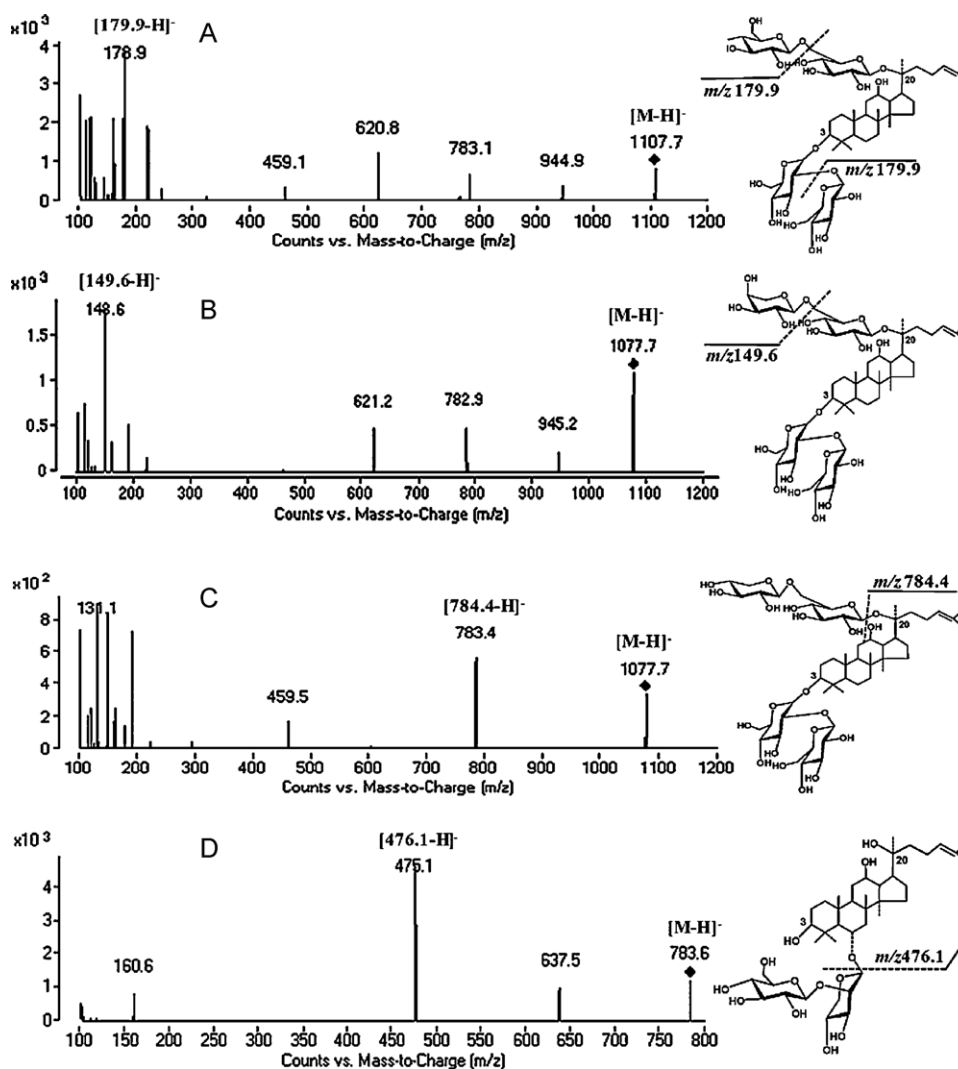


Fig. 1. Chemical structures and product ion mass spectra of Rb<sub>1</sub> (A), Rb<sub>2</sub> (B), Rb<sub>3</sub> (C) and IS (D).

ginsenosides and to systematically compare the similarities and differences in pharmacokinetic behaviors of the three ginsenosides.

## 2. Materials and methods

### 2.1. Chemicals and reagents

Ginsenoside Rb<sub>1</sub>, Rb<sub>2</sub>, Rb<sub>3</sub>, and Rg<sub>2</sub> (internal standard, IS) (99.0%) were obtained from the College of Chemistry, Jilin University (Changchun, China). Ammonium formate was obtained from Agilent Technologies. All other reagents were of HPLC grade.

### 2.2. Instrumentation and analytical conditions

The Agilent 1200 rapid resolution HPLC system was interfaced with an Agilent 6410 triple quadrupole mass spectrometer equipped with an electrospray ionization (ESI). The column was an Agilent Zorbax SB-C18 column (50 mm × 4.6 mm, 1.8 μm). The mobile phase was methanol and 1 mM ammonium formate (74:26, v/v, pH 6.0), the flow rate was 0.4 ml/min. Quantitation was performed in negative multiple reaction monitoring of  $m/z$  1107.7 →  $m/z$  178.9,  $m/z$  1077.7 →  $m/z$  148.6,  $m/z$  1077.7 →  $m/z$  783.4, and  $m/z$  783.6 →  $m/z$  475.1 for Rb<sub>1</sub>, Rb<sub>2</sub>, Rb<sub>3</sub>, and IS, respectively. The values of fragmentation/collision energy were

at 250 V/55 V, 210 V/67 V, 200 V/58 V for Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub>, respectively. Flow rate, 9 l/min; nebulizer pressure, 40 psi; gas temperature, 350 °C.

### 2.3. Preparation of calibration curve and plasma samples

Calibration curve were prepared by spiking blank rat plasma with the stock solution to the concentrations: 20, 50, 100, 500, 1000 ng/ml for Rb<sub>1</sub> and Rb<sub>2</sub>; 50, 100, 500, 1000, and 2500 ng/ml for Rb<sub>3</sub>. The sample (100 μl) was mixed with the IS solution (1 μg/ml, 10 μl) and extracted with 900 μl saturated N-butanol. After centrifugation at 13,000 × g for 10 min, the supernatant was evaporated to dryness under a stream of nitrogen. The residue was dissolved in 100 μl mobile phase and centrifuged for 10 min. 20 μl of the supernatant was injected into the LC-MS/MS.

### 2.4. Method validation

The matrix effect was evaluated by comparing the response of analyte spiked after extraction to that of an equivalent concentration of the standard solution. QC samples were prepared at three concentrations: 50, 100, and 1000 ng/ml for Rb<sub>1</sub> and Rb<sub>2</sub>; 100, 500, 2500 ng/ml for Rb<sub>3</sub>. The intra- and inter-day assays were evaluated of QC samples (n = 5) in rat plasma on three independent days. The extraction recovery was determined by comparing the response of

**Table 1**  
Pharmacokinetic parameters of Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub> after i.v. (10 mg/kg) and oral (50 mg/kg) administration in rats (n = 6).

Pharmacokinetic parameters	Rb <sub>1</sub>		Rb <sub>2</sub>		Rb <sub>3</sub>	
	i.v.	p.o.	i.v.	p.o.	i.v.	p.o.
AUC <sub>0-t</sub> (mg h/l)	1627.5 ± 138.9	63.5 ± 36.3	1642.7 ± 222.1	6.4 ± 1.9	1437.0 ± 108.6	37.4 ± 20.2
AUC <sub>0-∞</sub> (mg h/l)	1859.1 ± 190.0	66.8 ± 34.0	2006.3 ± 283.0	9.7 ± 3.2	2229.9 ± 684.6	55.1 ± 29.4
t <sub>1/2</sub> (h)	12.5 ± 1.8	9.8 ± 6.9	15.4 ± 3.7	23.1 ± 3.7	24.9 ± 12.6	21.1 ± 9.8
MRT <sub>0-t</sub> (h)	11.9 ± 0.7	9.7 ± 2.5	12.7 ± 0.9	14.4 ± 1.3	13.4 ± 0.5	13.9 ± 3.8
T <sub>max</sub> (h)	0.083	2.0 ± 0.9	0.083	4.8 ± 3.5	0.083	1.5 ± 2.0
C <sub>max</sub> (mg/l)	199.6 ± 12.4	6.1 ± 3.4	198.1 ± 27.9	0.4 ± 0.1	160.9 ± 31.6	3.3 ± 1.9

the analyte from the QC samples (n = 5) with the analyte spiked in post-extracted blank rat plasma at the same concentrations. QC samples (n = 3) were determined on the bench for 8 h at room temperature, three freeze–thaw cycles and processed-sample stability.

### 2.5. Pharmacokinetics study in rats

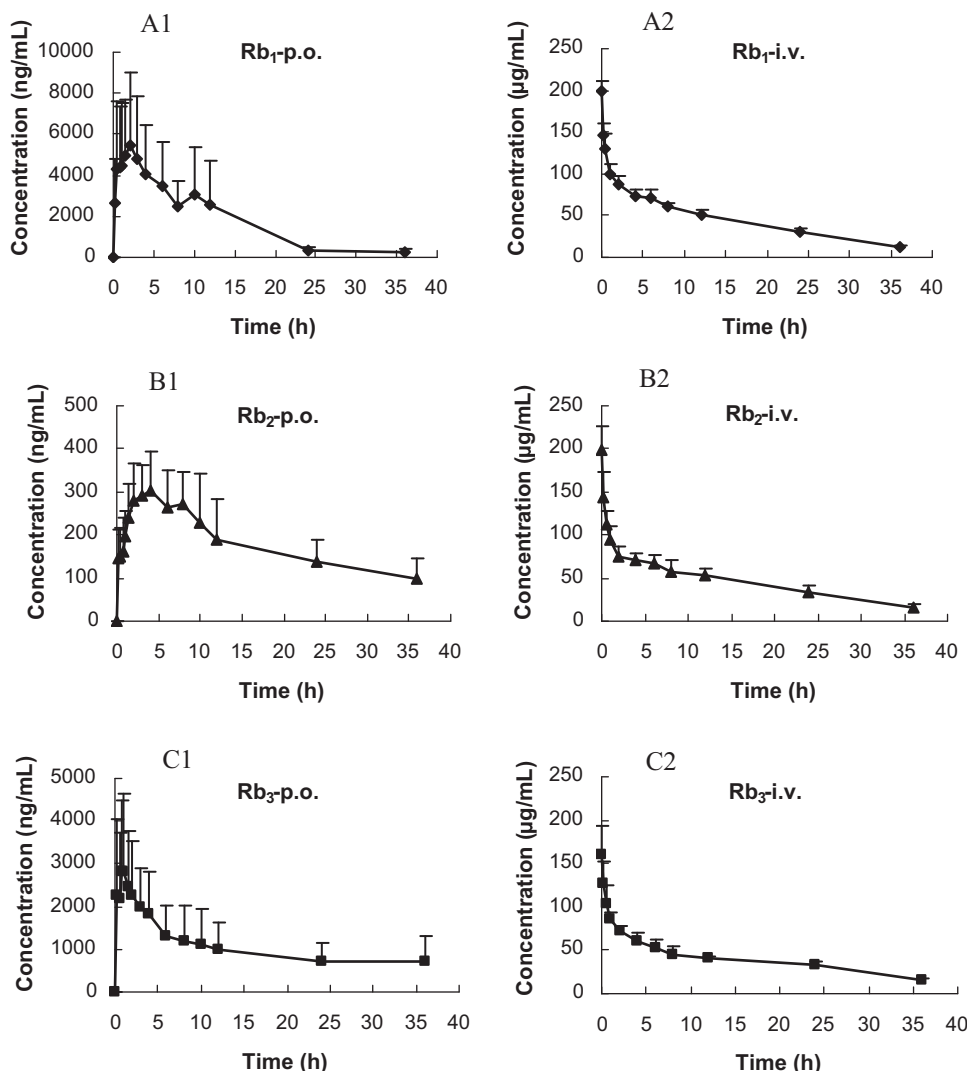
Sprague Dawley rats (200–220 g) were provided by Experimental Animal Center of Guangdong Province (Guangzhou, China). All studies were approved by the Ethics Committee of Southern Medical University. The rats were randomly assigned into six groups: three groups were intravenous administered of Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub>, respectively through tail vein (10 mg/kg), the three other

groups were orally administered of Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub>, respectively (50 mg/kg). Blood samples were collected at 0.083, 0.25, 0.5, 1, 2, 4, 6, 8, 12, 24, and 36 h (i.v.) and 0.25, 0.5, 0.75, 1, 1.5, 2, 3, 4, 6, 8, 10, 12, 24, and 36 h (p.o.). Blood samples were centrifuged immediately and stored at –20 °C until analysis.

## 3. Results and discussion

### 3.1. Method validation

The calibration curve was linear over the concentration range of 20–1000 ng/ml for Rb<sub>1</sub> and Rb<sub>2</sub>, and 50–2500 ng/ml for Rb<sub>3</sub>. The limits of detection (LOD) were 3.0 ng/ml, 4.0 ng/ml, and 6.5 ng/ml.



**Fig. 2.** Plasma concentration–time curve of Rb<sub>1</sub> (A), Rb<sub>2</sub> (B) and Rb<sub>3</sub> (C) after oral dose of 50 mg/kg and i.v. dose of 10 mg/kg (n = 6).

The results of matrix effect were in the range of 90–112% ( $n=5$ ). The intra- and inter-day accuracy were within 86–114% and the precision were within acceptable limits at three concentrations ( $n=5$ ). The extraction recovery was within 52–117%. All samples in stability evaluated displayed variability of less than 15% (R.S.D.).

### 3.2. Application to pharmacokinetics study

The method was successfully applied to pharmacokinetic study of Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub> in rats. The pharmacokinetic parameters (Table 1) were calculated by the non-compartmental mode using DAS2.0 software. The plasma concentrations versus time profiles are presented in Fig. 2. The results of  $T_{max}$  indicated that the absorption of Rb<sub>3</sub> is the fastest and Rb<sub>1</sub> is the slowest. The  $t_{1/2}$  after intravenous administration indicates that the three ginsenosides are eliminated slowly in vivo. Comparatively, the elimination of Rb<sub>1</sub> is the fastest, while Rb<sub>3</sub> is the slowest. The  $AUC_{0-36h}$  value for Rb<sub>1</sub> following oral administration was about 10 times larger than that of Rb<sub>2</sub> and was nearly two times larger than that of Rb<sub>3</sub>, suggesting that the absorption of Rb<sub>1</sub> in rats is much higher than Rb<sub>2</sub> and Rb<sub>3</sub>. The bioavailability was 0.78% for Rb<sub>1</sub>, 0.08% for Rb<sub>2</sub>, and 0.52% for Rb<sub>3</sub>, all the three ginsenosides are poor oral absorption. Since the pharmacokinetic characteristics of the three ginsenosides have been compared systematically, we presume that the pharmacokinetic behaviors of ginsenosides have structural preferences. In particular, ginsenosides with hexose and hydroxyl groups (Rb<sub>1</sub>) could present better oral absorption than those with pentose groups in the same glycosylation site.

### 4. Conclusions

A simple, rapid, and sensitive LC–MS/MS method for determination of ginsenoside in rat plasma was developed. The assay was successfully applied to pharmacokinetic study of Rb<sub>1</sub>, Rb<sub>2</sub>, and Rb<sub>3</sub> in rats. All three ginsenosides had poor bioavailability. The pharmacokinetic behaviors of ginsenosides have structural preference.

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### References

- [1] H.C. Ren, J.G. Sun, G.J. Wang, J.Y. A. H.T. Xie, W.B. Zha, B. Yan, F.Z. Sun, H.P. Hao, S.H. Gu, L.S. Sheng, F. Shao, J. Shi, F. Zhou, Sensitive determination of 20(S)-protopanaxadiol in rat plasma using HPLC–APCI–MS: application of pharmacokinetic study in rats, *J. Pharm. Biomed. Anal.* 48 (2008) 1476–1480.
- [2] M. Liu, H. Wang, S. Zhao, X. Shi, Y. Zhang, H. Xu, Y. Wang, X. Li, L. Zhang, Studies on target tissue distribution of ginsenosides and epimedium flavonoids in rats after intravenous administration of Jiweiling freeze-dried powder, *Biomed. Chromatogr.* 25 (2011) 1260–1272.
- [3] M. Song, S. Zhang, X. Xu, T. Hang, L. Jia, Simultaneous determination of three *Panax notoginseng* saponins at sub-nanograms by LC–MS/MS in dog plasma for pharmacokinetics of compound Danshen tablets, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.* 878 (2010) 3331–3337.
- [4] W. Chen, Y. Dang, C. Zhu, Simultaneous determination of three major bioactive saponins of *Panax notoginseng* using liquid chromatography–tandem mass spectrometry and a pharmacokinetic study, *Chin. Med.* 5 (2010) 12–17.
- [5] Q.F. Xu, X.L. Fang, D.F. Chen, Pharmacokinetics and bioavailability of ginsenoside Rb<sub>1</sub> and Rg<sub>1</sub> from *Panax notoginseng* in rats, *J. Ethnopharmacol.* 84 (2003) 187–192.
- [6] H. Liu, J. Yang, F. Du, X. Gao, X. Ma, Y. Huang, F. Xu, W. Niu, F. Wang, Y. Mao, Y. Sun, T. Lu, C. Liu, B. Zhang, C. Li, Absorption and disposition of ginsenosides after oral administration of *Panax notoginseng* extract to rats, *Drug Metab. Dispos.* 37 (2009) 2290–2298.
- [7] G.F. Deng, D.L. Wang, M.X. Meng, F. Hu, T.W. Yao, Simultaneous determination of notoginsenoside R<sub>1</sub>, ginsenoside Rg<sub>1</sub>, Re, Rb<sub>1</sub> and icariin in rat plasma by ultra-performance liquid chromatography–tandem mass spectrometry, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.* 877 (2009) 2113–2122.
- [8] Y.W. Ha, K.S. Ahn, J.C. Lee, S.H. Kim, B.C. Chung, M.H. Choi, Validated quantification for selective cellular uptake of ginsenosides on MCF-7 human breast cancer cells by liquid chromatography–mass spectrometry, *Anal. Bioanal. Chem.* 396 (2010) 3017–3025.
- [9] J. Sun, G. Wang, X. Haitang, L. Hao, P. Guoyu, I. Tucker, Simultaneous rapid quantification of ginsenoside Rg<sub>1</sub> and its secondary glycoside Rh<sub>1</sub> and aglycone protopanaxatriol in rat plasma by liquid chromatography–mass spectrometry after solid-phase extraction, *J. Pharm. Biomed. Anal.* 38 (2005) 126–132.
- [10] L. Liu, J. Huang, X. Hu, K. Li, C. Sun, Simultaneous determination of ginsenoside (G-Re, G-Rg<sub>1</sub>, G-Rg<sub>2</sub>, G-F1, G-Rh<sub>1</sub>) and protopanaxatriol in human plasma and urine by LC–MS/MS and its application in a pharmacokinetics study of G-Re in volunteers, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.* 879 (2011) 2011–2017.
- [11] H. Feng, W. Chen, C. Zhu, Pharmacokinetics study of bio-adhesive tablet of *Panax notoginseng* saponins, *Int. Arch. Med.* 4 (2011) 18–25.
- [12] H. Jin, J.H. Seo, Y.K. Uhm, C.Y. Jung, S.K. Lee, S.V. Yim, Pharmacokinetic comparison of ginsenoside metabolite IH-901 from fermented and non-fermented ginseng in healthy Korean volunteers, *J. Ethnopharmacol.* 139 (2011) 664–667.